

Human TGF-BETA 3 AccuSignal™ ELISA Kit - KOA0628

AccuSignal high sensitivity sandwich ELISA (Enzyme-Linked Immunosorbent Assay) Kit is *an in vitro* enzyme-linked immunosorbent assay designed for the quantitative detection of human TGF-BETA 3 in cell culture supernatants, serum, plasma (EDTA) and urine.

I. KIT COMPONENTS

Human TGF-BETA 3 AccuSignal™ ELISA Kit				
Component	Catalog Number	Description	Size	Storage of opened/reconstituted material
Antibody Coated ELISA Plate	KOA0628A	96-well plate pre-coated with anti-human TGF-BETA 3 antibody	1	Return unused wells to the foil pouch. Reseal along the entire edge of the zip-seal. May be stored for up to 1 month at 4°C provided this is within the expiration date of the kit.
Target Protein Standard	KOA0628B	lyophilized recombinant human TGF-BETA 3	2 vials (10 ng/vial)	Discard the TGFB3 stock solution after 12 hours at 4°C. May be stored at -20°C for 48 hours.
Biotinylated Detection Antibody	KOA0628C	biotinylated anti-human TGF-BETA 3 antibody	100 µl (dilution 1:100)	May be stored for up to 1 month at 4°C provided this is within the expiration date of the kit.
ABC Complex	KAB0101	Avidin-Biotin-Peroxidase Complex (ABC)	100 µl (dilution 1:100)	
Sample Diluent	KAE0101	sample diluent buffer	30 ml	
Antibody Diluent	KAF0101	antibody diluent buffer	12 ml	
ABC Diluent	KAG0101	ABC diluent buffer	12 ml	
Substrate Solution	KAC0101	TMB color developing reagent	10 ml	
Stop Solution	KAD0101	TMB stop solution	10 ml	
Adhesive Cover	-	Adhesive cover	4	

II. STORAGE CONDITIONS

Store at 4°C for 6 months, or at -20°C for 12 months from date of manufacture. Avoid multiple freeze-thaw cycles.

III. INTRODUCTION

Human TGF-BETA 3 ELISA Kit is based on standard sandwich enzyme-linked immune-sorbent assay technology. This assay employs a monoclonal antibody specific for TGF-BETA 3 pre-coated onto 96-well plate. Standards (Expression system for standard: NSO; Immunogen sequence: A301-S412) and test samples are pipetted into the wells, and a biotinylated detection polyclonal antibody from goat specific for TGF-BETA 3 is added to the wells. The wells are washed with PBS or TBS buffer. After washing away unbound biotinylated antibody, Avidin-Biotin-Peroxidase Complex is added to the wells and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB solution is added to the wells to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product. The addition of stop solution changes the color from blue to yellow. The density of yellow is proportional to the amount of Human TGF-BETA 3 amount captured in plate.

IV. REQUIRED EQUIPMENT AND REAGENTS

- Microplate reader in standard size
- Automated plate washer
- Adjustable pipettes and pipette tips
- Clean tubes and Eppendorf tubes
- Washing buffer (neutral PBS or TBS)
 - Preparation of 0.01M TBS:
Add 1.2g Tris, 8.5g NaCl; 450µl of purified acetic acid or 700µl of concentrated hydrochloric acid to 1000 ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.
 - Preparation of 0.01 M PBS:
Add 8.5 g sodium chloride, 1.4 g Na₂HPO₄ and 0.2 g NaH₂PO₄ to 1000 ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.

V. GENERAL CONSIDERATIONS

Please read the following instructions before starting the experiment.

- To inspect the validity of experimental operation and the appropriateness of sample dilution proportions, pilot experiment using standards and a small number of samples is recommended.
- The TMB Color Developing agent is colorless and transparent before using, contact us if it is not the case.
- Before using the Kit, briefly spin down the vials.
- For statistical reasons, we recommend both standard and samples should be assayed with a minimum of two replicates (duplicates).
- Do not let 96-well plate to dry, this will inactivate active components on plate.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.

- Do not mix or substitute reagents or materials from other kit lots or vendors.
- In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37°C for 30 min before using.

VI. SANDWICH ELISA PROTOCOL

A. Sample Preparation

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

- **Serum:**
Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 min. Analyze the serum immediately or aliquot and store at -20°C.
- **Cell culture supernatants:**
Remove particulates by centrifugation, assay immediately or aliquot and store at -20°C.
- **Plasma:**
Collect plasma using EDTA as an anticoagulant. Centrifuge for 15 min at 1000 x g within 30 min of collection. Assay immediately or aliquot and store at -20°C.
 - **Note:** it is important not to use any anticoagulant other than those listed above to treat plasma. Other anticoagulants could block the antibody binding site.
- **Urine:**
Collect the first urine of the day, micturate directly into a sterile container. Remove impurities by centrifugation. Assay immediately or aliquot and store samples at -20°C.
 - **Note:**
To detect TGFB3 in samples, you need to activate TGFB3 in samples prior to the assay. TGF beta 3 is mostly contained as inactive form in samples, please activate it before assay. It is unnecessary to activate the recombinant TGF-beta 3.
 - Solution A: 1N HCl: add 8.33ml of 12N HCl into 91.67ml of H₂O.
 - Solution B: 1.2N NaOH/0.5M HEPES: add 12ml of 10N NaOH and 11.9g HEPES into 75ml of H₂O, add H₂O to adjust volume to 100ml.
 - **Activate the sample**
 - **Cell culture supernates, urine:** add activating reagent pro rata, i.e. add 20µl of Solution A into 100µl of sample, 10 min later, add 20µl of Solution B. PH 7.0-7.6.
 - **Serum, plasma(EDTA):** add activating reagent pro rata, i.e. add 20µl of Solution A into 40µl of sample, 10 min later, add 20µl of Solution B. PH 7.0-7.6.
 - **Note:** Sample was diluted partly after adding activating reagent, so please pay attention to this when calculate target protein concentration.

B. Sample Dilution

The target protein concentration should be estimated, and appropriate sample dilutions should be selected such that the final protein concentration lies near the middle of the linear dynamic range of the assay. It is recommended to prepare 150 μ L of sample for each replicate to be assayed. The samples should be diluted with sample diluent and mixed gently.

C. Reagent Preparation

Reconstitution of the Human TGF-BETA 3 standard:

TGF-BETA 3 standard solution should be prepared no more than 2 hours prior to the experiment. Two vials of TGF-BETA 3 standard (10ng/vial) are provided with each kit. Use one vial/experiment.

- For preparation of 10,000pg/ml of Human TGF-BETA 3 standard solution, add 1ml sample diluent buffer into one vial, keep the vial at room temperature for 10 min and mix thoroughly.
- Label tubes 1-8. Tube # 1 –2000pg/ml, #2 –1000pg/ml, #3 – 500pg/ml, #4 – 250pg/ml, #5 – 125pg/ml, #6 – 62.5pg/ml, #7 – 31.25pg/ml, #8 – 0.0 (Blank).
- To generate standard #1, pipette 200 μ L of reconstituted 10ng/mL stock standard into tube#1, add 800 μ L sample diluent buffer and mix thoroughly.
- Pipette 300 μ L of the sample diluent buffer into tubes 2-7.
- Transfer 300 μ L from tube #1 to tube #2 and mix thoroughly.
- Transfer 300 μ L from tube #2 to tube #3 and mix thoroughly, continue further serial dilutions through tube #7.
- Tube#8 is left blank.

Note: The standard solutions are best used within 2 hours. The 10,000pg/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

Preparation of biotinylated anti-Human TGF-BETA 3 antibody working solution:

The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be 0.1ml/well x (the number of wells). Prepare 100-200 μ l more of the solution than the total volume required to compensate for pipetting errors.
- Biotinylated anti-Human TGF-BETA 3 antibody should be diluted 1:100 with the antibody diluent buffer and mixed thoroughly. For example, add 1 μ l Biotinylated Anti-Human TGF-BETA 3 antibody to 99 μ l antibody diluent buffer.

Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution:

The solution should be prepared no more than 1 hour prior to the experiment.

- The total volume should be: 0.1 ml/well x (the number of wells). Prepare 0.1-0.2 ml more of the solution than total volume required.
- Avidin-Biotin-Peroxidase Complex (ABC) should be diluted 1:100 with the ABC dilution buffer and mixed thoroughly. For example, add 1 μ l ABC to 99 μ l ABC diluent buffer.

D. Assay Procedure

ABC working solution and TMB substrate reagent should be warmed at 37°C for 30 min prior to use. Samples and reagents should be mixed thoroughly and evenly after dilution. A standard curve for TGF-BETA 3 detection should be prepared for each experiment. It is recommended to determine sample fold dilution by simple estimation of TGF-BETA 3 amount in the samples.

1. Aliquot 100µl per well of each human TGF-BETA 3 standard (2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, 62.5pg/ml, 31.25pg/ml) into pre-coated 96-well plate. Add 100µl of the sample diluent buffer into control wells. Add 100µl of diluted sample (cell culture supernatants, serum, plasma {EDTA} or urine into appropriate wells). See "Sample Dilution" above for details. **It is recommended to assay all standards, controls and samples in duplicate.**
2. Seal the plate with provided adhesive plastic cover and incubate at 37°C for 90 min.
3. Remove the cover and discard the solution. Invert the plate and blot it against clean paper towels or other absorbent material. Do NOT let the wells dry completely at any time.
4. Add 100µl of biotinylated anti-Human TGF-BETA 3 antibody working solution to each well, seal the plate with adhesive plastic cover and incubate at 37°C for 60 min.
5. Discard the solution and wash the plate 3 times with 0.01M TBS or 0.01M PBS (wash buffer) and each time let washing buffer stay in the wells for ~1 min. Wash by filling each well with 300µl of wash buffer using a multi-channel pipette or auto washer. Complete removal of liquid at each step is essential to good performance. (Note: for automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling the wells with PBS or TBS buffer). After the last wash, remove any remaining wash buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels or other absorbent material.
6. Add 100µl of prepared ABC working solution to each well, seal the plate and incubate at 37°C for 30 min.
7. Wash plate 5 times with 0.01M TBS or 0.01M PBS as described in step 5.
8. Add 90µl of TMB substrate reagent to each well, seal the plate and incubate for 25-30 min at 37°C in the dark. (Note: the optimal incubation time should be determined by the end user).
9. Add 100µl of stop solution to each well. The color will change to yellow immediately.
10. Read at 450 nm in a microplate reader within 30 min after adding the stop solution.

Calculations

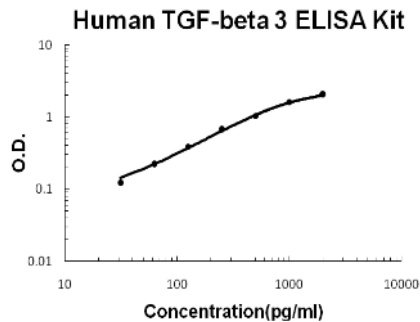
For calculations, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The Human TGF-BETA 3 concentration of the samples can be interpolated from the standard curve.

Note: if the samples measured were diluted, multiply the concentrations obtained from interpolation by the dilution factor.

VII. DATA OBTAINED FROM HUMAN TGF-BETA 3

Concentration (pg/ml)	0	31.2	62.5	125	250	500	1000	2000
O.D	0.031	0.133	0.180	0.316	0.534	0.913	0.913	1.813

(TMB reaction incubation at 37°C for 25-30min)



VIII. HUMAN TGF-BETA 3 ACCUSIGNAL™ ELISA KIT STANDARD CURVE

This standard curve was generated for demonstration purpose only
 (A standard curve must be run with each assay).

Range	31.2pg/ml-2000pg/ml
Sensitivity	<10pg/ml
Specificity	Natural and recombinant Human TGF-BETA 3
Cross-reactivity	There is no detectable cross-reactivity with other relevant proteins.

IX. INTRA AND INTER ASSAY PRECISION

Intra-Assay Precision (Precision within an assay): Three samples of known concentration were tested on one plate to assess intra-assay precision.

Inter-Assay Precision (Precision between assays): Three samples of known concentration were tested in separate assays to assess inter-assay precision.

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
N	16	16	16	24	24	24
Mean(pg/ml)	66	244	776	70	242	733
Standard deviation	2.83	13.17	35.49	3.71	14.27	42.51
CV (%)	4.3	5.4	4.6	5.3	5.9	5.8

X. ASSAY SUMMARY

- Prepare all reagents, samples and standards as instructed.
- Add standards or samples to each well used. Incubate the plate at 37°C for 90 min. Do not wash.
- Add prepared biotinylated antibody to each well. Incubate the plate at 37°C for 60 min. Wash plate 3 times with 0.01M TBS or PBS.
- Add prepared ABC working solution. Incubate the plate at 37°C for 30 min. Wash plate 5 times with 0.01M TBS or TBS.
- Add TMB developing reagent to each well. Incubate the plate at 37°C in dark for 25-30 min.
- Add TMB stop solution. Read at 450nm.

XI. TROUBLESHOOTING

Weak or No Signal

Possible Cause	Solution
Problem with the standard	Use new sample Check that the standard is appropriately handled
Incubation time too short	Follow the exact guidelines for incubation time (If the problem persists, try incubating samples at 4°C overnight)
Incubation temperature too low	Ensure incubations are done at correct temperature Before proceeding
Incompatible sample type	Use sample that the assay is known to detect as a positive control (Include such control in your experiment)
Incompatible assay buffer	Ensure assay buffer is compatible with the target of interest
Target present below detection limit	Decrease dilution factor or concentrate samples
Incorrect/Insufficient/No substrate	Check the substrate identity Increase concentration or amount of substrate
Antibody stored at 4°C for several weeks or subjected to repeat freeze-thaw cycles	Use fresh aliquot of antibody that has been stored at -20°C or below
Incorrect reagents added/prepared; Missing reagents	Check protocol, ensure correct reagents are added in proper order and prepared to correct concentrations
Expired/Contaminated reagents	Prepare fresh/uncontaminated reagents
Enzyme inhibitor present	Avoid sodium azide in HRP reactions
Incorrect storage of components	Check storage conditions for the kit (Kit need to be stored at 4°C)
Excessive plate washing	Gently pipette wash buffer (manual method) Ensure correct pressure (automatic wash system)
Wells dry out	Cover plate using adhesive cover at all incubation times
Plate read at incorrect detection wavelength	Use recommended wavelength/filter Ensure plate reader is set correctly for substrate used

Slow color development	Prepare substrate immediately before use Allow longer incubation time Ensure stock solution is unexpired and uncontaminated
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Saturated Signal

Possible Cause	Solution
High sample concentration	Use higher sample dilutions (Determine the optimal dilutions by titration assay)
Excessive substrate	Decrease concentration or amount of substrate: The substrate provided with the ELISA kit might require further dilution
Substrate color changed before use	Prepare substrate immediately before use
Non-specific antibody binding	Use affinity-purified antibody and preferably one that is pre-adsorbed.
Incubation time too long	Follow the exact guidelines (If the problem persists, try incubating samples at 4°C overnight)
Excessive antibody	Repeat the assay with lower antibody concentrations to find the optimal one for your experiment
Contaminated buffers or HRP	Prepare and use fresh buffers
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and dry the plate on a paper towel
Plate adhesive cover not used or re-used	During incubations, cover plates with adhesive cover. Use a fresh cover every time the used cover is removed from the plate
Plate read at incorrect wavelength	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used
Excess time before plate reading	Read your plate within 30 minutes after adding the substrate (If the reading is not performed within this time frame, add a stopping solution after sufficient color is developed in the plate)

High Background

Possible Cause	Solution
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and pat dry the plate on a paper towel
Excessive antibody	Repeat the assay with lower antibody concentrations to find the optimal one for your experiment
Excessive substrate	Decrease concentration or amount of substrate
Cross reactivity	Run appropriate controls
Non-specific antibody binding	Use affinity-purified antibody and preferably one that is pre-adsorbed
Insufficient Tween in buffers	Use PBS or TBS containing 0.05% Tween
Suboptimal salt concentration in washing buffer	Optimize salt concentration as high concentration can reduce non-specific interactions
Incubation temperature too high	Optimize incubation temperature for your assay (antibodies bind optimally at very specific temperature)

Reagents were not mixed properly	Thoroughly mix all reagents and samples before pipetting solutions into wells
Blanks contaminated with samples	Change pipette tips when switching between blanks and samples
Sample contaminated with enzymes	Test samples with substrate alone to check for contaminating enzymes
Contaminated TMB substrate	Use a clean container to check that the substrate is not contaminated (TMB substrate should be clear and colorless before adding to wells)
Substrate exposed to light	Carry out substrate incubation in dark
Evaporation of solution from well during incubation	Always incubate with a cover on the plate
Incubation time too long	Follow the exact guidelines for incubation times (If the problem persists, try incubating samples at 4°C overnight)
Incorrect standard curve dilutions	Check pipetting techniques Check calculations
Unstopped color development	Use Stopping solution to prevent over-development
Excessive time lapsed before plate reading	Read your plate within 30 minutes after adding the substrate (If the reading is not performed within this time frame, add a stopping solution after sufficient color is developed in the plate)
Incorrect plate reading setting	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used

Low Sensitivity

Possible Cause	Solution
Improper storage of ELISA kit	Store all reagents as recommended
Insufficient target	Reduce sample dilution or concentrate sample
Inactive substrate	Ensure reporter enzyme has the expected activity
Insufficient substrate	Increase concentration or amount of substrate
Incompatible sample type	Include positive control in your experiment
Interfering ingredients in buffers and sample	Check reagents for any interfering chemicals, e.g. sodium azide in antibodies inhibit HRP enzyme; EDTA used as anti-coagulant for plasma collection inhibits enzymatic reactions
Mixing or substituting reagents from different kits	Avoid mixing components from different kits
Incorrect plate reading setting	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used

Poor Standard Curve Generation

Possible Cause	Solution
Improper standard solution	Confirm dilutions are done correctly Prepare new standard curve as appropriate
Standard improperly reconstituted	Briefly spin vial before opening Inspect for undissolved material after reconstituting
Standard degraded	Store and handle standard as recommended Prepare standards no more than two hours before use

Pipetting error	Use calibrated pipettes and proper pipetting technique
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and pat dry the plate on a paper towel
Poorly mixed reagents	Thoroughly mix reagents
Plates stacked during incubation	Keep plates separated if not using rotating plates

Poor Replicate Data

Possible Cause	Solution
Bubbles in wells	Ensure no bubbles are present prior to reading the plate
Insufficient washing of wells	Carefully wash wells Check that all ports of the plate washer are unobstructed
Incomplete reagent mixing	Ensure all reagents are mixed thoroughly
Inconsistent pipetting	Use calibrated pipettes and proper pipetting techniques Use a new cover every time the used cover is removed from the plate
Inconsistent sample preparation or storage	Ensure consistent sample preparation and optimal sample storage (e.g. minimize freeze/thaw cycles)
Particulates in samples	Remove the particulates by centrifugation
Cross-well contamination	Ensure plate covers and pipette tips are not contaminated with reagents
Edge effect (higher or lower OD in peripheral wells than in central wells)	Ensure plates and reagents are kept at temperatures as instructed During incubation, seal the plate completely and avoid stacking plates

Inconsistent Assay-to-Assay Results

Possible Cause	Solution
Insufficient washing of wells	Carefully wash wells Check that all ports of the plate washer are unobstructed
Varied incubation temperatures	Adhere to recommended incubation temperature
Variation in protocol	Adhere to the same protocol from experiment to experiment
Plate cover not used or re-used	During incubations, cover plates with plate cover Use a new cover every time the used one is removed
Incorrect dilutions	Confirm dilutions are done correctly for standard solutions Prepare new standard curve as appropriate
Contaminated buffers	Prepare and use fresh buffers

Slow Color Development

Possible Cause	Solution
Substrates too old, contaminated or used at incorrect pH	Prepare fresh substrates at correct pH
Expired/Contaminated solutions	Prepare fresh reagents before use

Incorrect incubation temperature	Ensure plates and reagents are kept at temperatures as instructed During incubation, seal the plate completely and avoid stacking plates
Low antibody concentration	Repeat the assay with higher antibody concentrations to find the optimal one for your experiment

Plate Imaging Problem

Possible Cause	Solution
Oversaturated image after acquisition	Use full resolution image to analyze results (Do not use jpeg or other compressed formats)
Blurry spots in images	Re-focus your camera before taking a new image
Repeated pixel values or rectangular spots	Use lower bin size, higher image resolution and/or lossless file type
Flat standard in images	Reduce acquisition time

XII. RELATED PRODUCTS

Component	Catalog #	Size
10x TBS pH 7.5	MB-012	1000 ml
10x TTBS pH 7.5	MB-013	1000 ml
10X PBS pH 7.2	MB-008	1000 ml
10x PBST pH 7.2	MB-075-1000	1000 ml