Mouse IGF-1 AccuSignal™ ELISA Kit – KOA0195

AccuSignal high sensitivity sandwich ELISA (Enzyme-Linked Immunosorbent Assay) Kit is *an in vitro* enzyme-linked immunosorbent assay designed for the quantitative detection of Mouse IGF-1 in cell culture supernatants, cell lysates, serum, plasma (heparin, EDTA).

I.   KIT COMPONENTS

Mouse IGF-1 AccuSignal™ ELISA Kit			
Component	Catalog Number	Description	Size
Antibody Coated ELISA Plate	KOA0195A	96-well plate pre-coated with anti-Mouse IGF-1 antibody	1
Target Protein Standard	KOA0195B	lyophilized recombinant Mouse IGF-1	2 vials (10 ng/vial)
Biotinylated Detection Antibody	KOA0195C	biotinylated anti-Mouse IGF-1 antibody	130 µl (dilution 1:100)
ABC Complex	KAB0101	Avidin-Biotin-Peroxidase Complex (ABC)	130 µl (dilution 1:100)
Sample Diluent	KAE0101	sample diluent buffer	30 ml
Antibody Diluent	KAF0101	antibody diluent buffer	12 ml
ABC Diluent	KAG0101	ABC diluent buffer	12 ml
Substrate Solution	KAC0101	TMB color developing reagent	10 ml
Stop Solution	KAD0101	TMB stop solution	10 ml
Adhesive Cover	-	Adhesive cover	4

II.   STORAGE CONDITIONS

Store at 4°C for 6 months, or at -20°C for 12 months from date of manufacture. Avoid multiple freeze-thaw cycles.

III. INTRODUCTION

Mouse IGF-1 ELISA Kit is based on standard sandwich enzyme-linked immune-sorbent assay technology. This assay employs a monoclonal antibody specific for IGF-1 pre-coated onto 96-well plate. Standards (Expression system for standard: E. coli; Immunogen sequence: G49-A118) and test samples are pipetted into the wells, and a biotinylated detection polyclonal antibody from goat specific for IGF-1 is added to the wells. The wells are washed with PBS or TBS buffer. After washing away unbound biotinylated antibody, Avidin-Biotin-Peroxidase Complex is added to the wells and unbound conjugates are washed away with PBS or TBS buffer. HRP substrate TMB solution is added to the wells to visualize HRP enzymatic reaction. TMB is catalyzed by HRP to produce a blue color product. The addition of stop solution changes the color from blue to yellow. The density of yellow is proportional to the amount of Mouse IGF-1 amount captured in plate.

IV. REQUIRED EQUIPMENT AND REAGENTS

- Microplate reader in standard size
- Automated plate washer
- Adjustable pipettes and pipette tips
- Clean tubes and Eppendorf tubes
- Washing buffer (neutral PBS or TBS)
  - Preparation of 0.01M TBS:
    Add 1.2g Tris, 8.5g NaCl; 450µl of purified acetic acid or 700µl of concentrated hydrochloric acid to 1000 ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.
  - Preparation of 0.01 M PBS:
    Add 8.5 g sodium chloride, 1.4 g Na_2HPO_4 and 0.2 g NaH_2PO_4 to 1000 ml distilled water and adjust pH to 7.2-7.6. Finally, adjust the total volume to 1L.

V. GENERAL CONSIDERATIONS

Please read the following instructions before starting the experiment.

- To inspect the validity of experimental operation and the appropriateness of sample dilution proportions, pilot experiment using standards and a small number of samples is recommended.
- The TMB Color Developing agent is colorless and transparent before using, contact us if it is not the case.
- Before using the Kit, briefly spin down the vials.
- For statistical reasons, we recommend both standard and samples should be assayed with a minimum of two replicates (duplicates).
- Do not let 96-well plate to dry, this will inactivate active components on plate.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Do not mix or substitute reagents or materials from other kit lots or vendors.
- In order to avoid marginal effect of plate incubation due to temperature difference (reaction may be stronger in the marginal wells), it is suggested that the diluted ABC and TMB solution be pre-warmed in 37℃ for 30 min before using.

VI. SANDWICH ELISA PROTOCOL

A. Sample Preparation

Store samples to be assayed within 24 hours at 2-8°C. For long-term storage, aliquot and freeze samples at -20°C. Avoid repeated freeze-thaw cycles.

- **Serum:**
  Allow the serum to clot in a serum separator tube (about 4 hours) at room temperature. Centrifuge at approximately 1000 X g for 15 min. Analyze the serum immediately or aliquot and store at -20°C.
- **Cell culture supernatants:**
  Remove particulates by centrifugation, assay immediately or aliquot and store at -20°C.

- **Plasma:**
  Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 min at 1000 x g within 30 min of collection. Assay immediately or aliquot and store at -20°C.
  - o **Note:** it is important not to use any anticoagulants other than those listed above to treat plasma. Other anticoagulants could block the antibody binding site.
- **Cell Lysates:**
  Lyse cells making sure there are no visible cell sediments. Centrifuge the cell lysates at approximately 10,000 x g for 5 minutes. Collect the supernatant.

B. Sample Dilution

Please note that levels of the target protein may vary between different samples. Estimation of the concentration of a target protein in the sample and the optimal dilution factors for each sample must be determined by the investigator. The diluted target protein concentration should fall near the middle of the linear regime in the standard curve. Sample diluent buffer should be used for dilution of samples.

Following are the suggested guidelines for sample dilution. **The samples must be mixed well with the diluent buffer**.

- **High target protein concentration (10,000pg/ml-100,000pg/ml)**
  The working dilution should be 1:100. For example, add 1μl sample into a tube with 99μl sample diluent buffer.
- **Medium target protein concentration (1000pg/ml-10,000pg/ml)**
  The working dilution should be 1:10. For example, add 10μl sample into a tube with 90μl sample diluent buffer.
- **Low target protein concentration (15.6pg/ml-1000pg/ml)**
  The working dilution should be 1:2. For example, add 50μl sample into a tube with 50μl sample diluent buffer.
- **Very Low target protein concentration (0pg/ml-15.6pg/ml)**
  No dilution necessary, or the working dilution should be 1:2.

C. Reagent Preparation

Reconstitution of the Mouse IGF-1 standard:

IGF-1 standard solution should be prepared no more than 2 hours prior to the experiment. Two vials of IGF-1 standard (10ng/vial) are provided with each kit. Use one vial/experiment.

- For preparation of 10,000pg/ml of Mouse IGF-1 standard solution, add 1ml sample diluent buffer into one vial, keep the vial at room temperature for 10 min and mix thoroughly.
- Label tubes 1-8. Tube # 1 –4000pg/ml, #2 –2000pg/ml, #3 – 1000pg/ml, #4 – 500pg/ml, #5 – 250pg/ml, #6 – 125pg/ml, #7 – 62.5pg/ml, #8 – 0.0 (Blank).
- To generate standard #1, pipette 400μL of reconstituted 10 ng/mL stock standard into tube#1, add 600μL of sample diluent buffer and mix thoroughly.
- Pipette 300μL of the sample diluent buffer into tubes 2-7.
- Transfer 300μL from tube #1 to tube #2 and mix thoroughly.
- Transfer 300μL from tube #2 to tube #3 and mix thoroughly, continue further serial dilutions through tube#7.
- Tube#8 is left blank.

Note: The standard solutions are best used within 2 hours. The 10,000pg/ml standard solution should be stored at 4°C for up to 12 hours, or at -20°C for up to 48 hours. Avoid repeated freeze-thaw cycles.

Preparation of biotinylated anti-Mouse IGF-1 antibody working solution:

The solution should be prepared no more than 2 hours prior to the experiment.

- The total volume should be 0.1ml/well x (the number of wells). Prepare 100-200µl more of the solution than the total volume required to compensate for pipetting errors.
- Biotinylated anti-Mouse IGF-1 antibody should be diluted 1:100 with the antibody diluent buffer and mixed thoroughly. For example, add 1µl Biotinylated Anti-Mouse IGF-1 antibody to 99µl antibody diluent buffer.

Preparation of Avidin-Biotin-Peroxidase Complex (ABC) working solution:

The solution should be prepared no more than 1 hour prior to the experiment.

- The total volume should be: 0.1 ml/well x (the number of wells). Prepare 0.1-0.2 ml more of the solution than total volume required.
- Avidin-Biotin-Peroxidase Complex (ABC) should be diluted 1:100 with the ABC dilution buffer and mixed thoroughly. For example, add 1µl ABC to 99µl ABC diluent buffer.

D. Assay Procedure

ABC working solution and TMB substrate reagent should be warmed at 37°C for 30 min prior to use. Samples and reagents should be mixed thoroughly and evenly after dilution. A standard curve for IGF-1 detection should be prepared for each experiment. It is recommended to determine sample fold dilution by simple estimation of IGF-1 amount in the samples.

1. Aliquot 100µl per well of each Mouse IGF-1 standard (4000pg/ml, 2000pg/ml, 1000pg/ml, 500pg/ml, 250pg/ml, 125pg/ml, and 62.5pg/ml) into pre-coated 96-well plate. Add 100µl of the sample diluent buffer into control wells. Add 100µl of diluted sample (cell culture supernatants, serum, plasma {heparin, EDTA} or cell lysates into appropriate wells). See "Sample Dilution" above for details. **It is recommended to assay all standards, controls and samples in duplicate.**
2. Seal the plate with provided adhesive plastic cover and incubate at 37°C for 90 min.
3. Remove the cover and discard the solution. Invert the plate and blot it against clean paper towels or other absorbent material. Do NOT let the wells dry completely at any time.
4. Add 100µl of biotinylated anti-Mouse IGF-1 antibody working solution to each well, seal the plate with adhesive plastic cover and incubate at 37°C for 60 min.
5. Discard the solution and wash the plate 3 times with 0.01M TBS or 0.01M PBS (wash buffer) and each time let washing buffer stay in the wells for ˜1 min. Wash by filling each well with 300µl of wash buffer using a multi-channel pipette or auto washer. Complete removal of liquid at each step is essential to good performance. (Note: for automated washing, aspirate all wells and wash THREE times with PBS or TBS buffer, overfilling the wells with PBS or TBS buffer). After the last wash, remove any remaining wash buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels or other absorbent material.
6. Add 100µl of prepared ABC working solution to each well, seal the plate and incubate at 37°C for 30 min.
7. Wash plate 5 times with 0.01M TBS or 0.01M PBS as described in step 5.

8. Add 90µl of TMB substrate reagent to each well, seal the plate and incubate for 25-30 min at 37°C in the dark. (Note: the optimal incubation time should be determined by the end user).
9. Add 100µl of stop solution to each well. The color will change to yellow immediately.
10. Read at 450 nm in a microplate reader within 30 min after adding the stop solution.

Calculations

For calculations, (the relative O.D.450) = (the O.D.450 of each well) – (the O.D.450 of Zero well). The standard curve can be plotted as the relative O.D.450 of each standard solution (Y) vs. the respective concentration of the standard solution (X). The Mouse IGF-1 concentration of the samples can be interpolated from the standard curve.
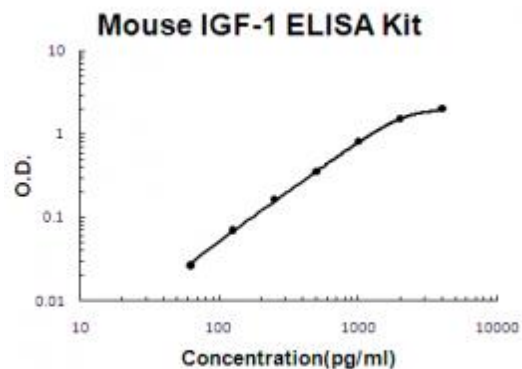
Note: if the samples measured were diluted, multiply the concentrations obtained from interpolation by the dilution factor.

VII. DATA OBTAINED FROM MOUSE IGF-1

Concentration (pg/ml)	0	62.5	125	250	500	1000	2000	4000
O.D	0.023	0.080	0.141	0.258	0.609	1.233	1.634	2.085

(TMB reaction incubation at 37°C for 25-30min)

VIII. MOUSE IGF-1 ACCUSIGNAL^{TM} ELISA KIT STANDARD CURVE

This standard curve was generated for demonstration purpose only (A standard curve must be run with each assay).

Range	62.5pg/ml-4000pg/ml
Sensitivity	<5pg/ml
Specificity	Natural and recombinant Mouse IGF-1
Cross-reactivity	There is no detectable cross-reactivity with IGF-2.

IX. INTRA AND INTER ASSAY PRECISION

Intra-Assay Precision (Precision within an assay): Three samples of known concentration were tested on one plate to assess intra-assay precision.

Inter-Assay Precision (Precision between assays): Three samples of known concentration were tested in separate assays to assess inter-assay precision.

	Intra-Assay Precision			Inter-Assay Precision		
Sample	1	2	3	1	2	3
N	16	16	16	24	24	24
Mean(pg/ml)	75	604	1350	69	617	1307
Standard deviation	5.85	24.76	93.15	6.69	30.23	92.79
CV (%)	7.8	4.1	6.9	9.7	4.9	7.1

X. ASSAY SUMMARY

- Prepare all reagents, samples and standards as instructed.
- Add standards or samples to each well used. Incubate the plate at 37°C for 90 min. Do not wash.
- Add prepared biotinylated antibody to each well. Incubate the plate at 37°C for 60 min. Wash plate 3 times with 0.01M TBS or PBS.
- Add prepared ABC working solution. Incubate the plate at 37°C for 30 min. Wash plate 5 times with 0.01M TBS or TBS.
- Add TMB developing reagent to each well. Incubate the plate at 37°C in dark for 25-30 min.
- Add TMB stop solution. Read at 450nm.

XI. TROUBLESHOOTING

Weak or No Signal

Possible Cause	Solution
Problem with the standard	Use new sample Check that the standard is appropriately handled
Incubation time too short	Follow the exact guidelines for incubation time (If the problem persists, try incubating samples at 4°C overnight)
Incubation temperature too low	Ensure incubations are done at correct temperature Before proceeding
Incompatible sample type	Use sample that the assay is known to detect as a positive control (Include such control in your experiment)
Incompatible assay buffer	Ensure assay buffer is compatible with the target of interest
Target present below detection limit	Decrease dilution factor or concentrate samples
Incorrect/Insufficient/No substrate	Check the substrate identity Increase concentration or amount of substrate

Antibody stored at 4°C for several weeks or subjected to repeat freeze-thaw cycles	Use fresh aliquot of antibody that has been stored at -20°C or below
Incorrect reagents added/ prepared; Missing reagents	Check protocol, ensure correct reagents are added in proper order and prepared to correct concentrations
Expired/Contaminated reagents	Prepare fresh/uncontaminated reagents
Enzyme inhibitor present	Avoid sodium azide in HRP reactions
Incorrect storage of components	Check storage conditions for the kit (Kit need to be stored at 4°C)
Excessive plate washing	Gently pipette wash buffer (manual method) Ensure correct pressure (automatic wash system)
Wells dry out	Cover plate using adhesive cover at all incubation times
Plate read at incorrect detection wavelength	Use recommended wavelength/filter Ensure plate reader is set correctly for substrate used
Slow color development	Prepare substrate immediately before use Allow longer incubation time Ensure stock solution is unexpired and uncontaminated

Saturated Signal

Possible Cause	Solution
High sample concentration	Use higher sample dilutions (Determine the optimal dilutions by titration assay)
Excessive substrate	Decrease concentration or amount of substrate: The substrate provided with the ELISA kit might require further dilution
Substrate color changed before use	Prepare substrate immediately before use
Non-specific antibody binding	Use affinity-purified antibody and preferably one that is pre-adsorbed.
Incubation time too long	Follow the exact guidelines (If the problem persists, try incubating samples at 4°C overnight)
Excessive antibody	Repeat the assay with lower antibody concentrations to find the optimal one for your experiment
Contaminated buffers or HRP	Prepare and use fresh buffers
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and dry the plate on a paper towel
Plate adhesive cover not used or re-used	During incubations, cover plates with adhesive cover. Use a fresh cover every time the used cover is removed from the plate
Plate read at incorrect wavelength	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used
Excess time before plate reading	Read your plate within 30 minutes after adding the substrate (If the reading is not performed within this time frame, add a stopping solution after sufficient color is developed in the plate)

High Background

Possible Cause	Solution
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and pat dry the plate on a paper towel

Excessive antibody	Repeat the assay with lower antibody concentrations to find the optimal one for your experiment
Excessive substrate	Decrease concentration or amount of substrate
Cross reactivity	Run appropriate controls
Non-specific antibody binding	Use affinity-purified antibody and preferably one that is pre-adsorbed
Insufficient Tween in buffers	Use PBS or TBS containing 0.05% Tween
Suboptimal salt concentration in washing buffer	Optimize salt concentration as high concentration can reduce non-specific interactions
Incubation temperature too high	Optimize incubation temperature for your assay (antibodies bind optimally at very specific temperature)
Reagents were not mixed properly	Thoroughly mix all reagents and samples before pipetting solutions into wells
Blanks contaminated with samples	Change pipette tips when switching between blanks and samples
Sample contaminated with enzymes	Test samples with substrate alone to check for contaminating enzymes
Contaminated TMB substrate	Use a clean container to check that the substrate in not contaminated (TMB substrate should be clear and colorless before adding to wells)
Substrate exposed to light	Carry out substrate incubation in dark
Evaporation of solution from well during incubation	Always incubate with a cover on the plate
Incubation time too long	Follow the exact guidelines for incubation times (If the problem persists, try incubating samples at 4°C overnight)
Incorrect standard curve dilutions	Check pipetting techniques Check calculations
Unstopped color development	Use Stopping solution to prevent over-development
Excessive time lapsed before plate reading	Read your plate within 30 minutes after adding the substrate (If the reading is not performed within this time frame, add a stopping solution after sufficient color is developed in the plate)
Incorrect plate reading setting	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used

Low Sensitivity

Possible Cause	Solution
Improper storage of ELISA kit	Store all reagents as recommended
Insufficient target	Reduce sample dilution or concentrate sample
Inactive substrate	Ensure reporter enzyme has the expected activity
Insufficient substrate	Increase concentration or amount of substrate
Incompatible sample type	Include positive control in your experiment
Interfering ingredients in buffers and sample	Check reagents for any interfering chemicals, e.g. sodium azide in antibodies inhibit HRP enzyme; EDTA used as anti-coagulant for plasma collection inhibits enzymatic reactions
Mixing or substituting reagents from different kits	Avoid mixing components from different kits
Incorrect plate reading setting	Use recommended wavelength/filter Ensure plate reader is set correctly for the substrate used

Poor Standard Curve Generation

Possible Cause	Solution
Improper standard solution	Confirm dilutions are done correctly Prepare new standard curve as appropriate
Standard improperly reconstituted	Briefly spin vial before opening Inspect for undissolved material after reconstituting
Standard degraded	Store and handle standard as recommended Prepare standards no more than two hours before use
Pipetting error	Use calibrated pipettes and proper pipetting technique
Insufficient washing	Follow the exact guidelines At the end of each washing step, flick the plate over a sink and pat dry the plate on a paper towel
Poorly mixed reagents	Thoroughly mix reagents
Plates stacked during incubation	Keep plates separated if not using rotating plates

Poor Replicate Data

Possible Cause	Solution
Bubbles in wells	Ensure no bubbles are present prior to reading the plate
Insufficient washing of wells	Carefully wash wells Check that all ports of the plate washer are unobstructed
Incomplete reagent mixing	Ensure all reagents are mixed thoroughly
Inconsistent pipetting	Use calibrated pipettes and proper pipetting techniques Use a new cover every time the used cover is removed from the plate
Inconsistent sample preparation or storage	Ensure consistent sample preparation and optimal sample storage (e.g. minimize freeze/thaw cycles)
Particulates in samples	Remove the particulates by centrifugation
Cross-well contamination	Ensure plate covers and pipette tips are not contaminated with reagents
Edge effect (higher or lower OD in peripheral wells than in central wells)	Ensure plates and reagents are kept at temperatures as instructed During incubation, seal the plate completely and avoid stacking plates

Inconsistent Assay-to-Assay Results

Possible Cause	Solution
Insufficient washing of wells	Carefully wash wells Check that all ports of the plate washer are unobstructed
Varied incubation temperatures	Adhere to recommended incubation temperature
Variation in protocol	Adhere to the same protocol from experiment to experiment
Plate cover not used or re-used	During incubations, cover plates with plate cover Use a new cover every time the used one is removed
Incorrect dilutions	Confirm dilutions are done correctly for standard solutions Prepare new standard curve as appropriate
Contaminated buffers	Prepare and use fresh buffers

Slow Color Development

Possible Cause	Solution
Substrates too old, contaminated or used at incorrect pH	Prepare fresh substrates at correct pH
Expired/Contaminated solutions	Prepare fresh reagents before use
Incorrect incubation temperature	Ensure plates and reagents are kept at temperatures as instructed During incubation, seal the plate completely and avoid stacking plates
Low antibody concentration	Repeat the assay with higher antibody concentrations to find the optimal one for your experiment

Plate Imaging Problem

Possible Cause	Solution
Oversaturated image after acquisition	Use full resolution image to analyze results (Do not use jpeg or other compressed formats)
Blurry spots in images	Re-focus your camera before taking a new image
Repeated pixel values or rectangular spots	Use lower bin size, higher image resolution and/or lossless file type
Flat standard in images	Reduce acquisition time

XII. RELATED PRODUCTS

Component	Catalog #	Size
10x TBS pH 7.5	MB-012	1000 ml
10x TTBS pH 7.5	MB-013	1000 ml
10X PBS pH 7.2	MB-008	1000 ml
10x PBST pH 7.2	MB-075-1000	1000 ml